

Resource Summary Report

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Fred Hutchinson Cancer Center Flow Cytometry Core Facility

RRID:SCR_022613

Type: Tool

Proper Citation

Fred Hutchinson Cancer Center Flow Cytometry Core Facility (RRID:SCR_022613)

Resource Information

URL: <https://www.fredhutch.org/en/research/shared-resources/core-facilities/flow-cytometry.html>

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Description: Includes services from sterile cell sorting for therapeutic applications to whole cell mass spectrometry, diagnostics and early stage clinical trials. Provides expertise from experimental design to data analysis and troubleshooting, therapeutic cell sorting. Supports isolation of cells for Phase 1 clinical trials of cellular immunotherapies.

Synonyms: Fred Hutchinson Cancer Center Flow Cytometry Shared Resource

Resource Type: core facility, access service resource, service resource

Keywords: sterile cell sorting, whole cell mass spectrometry, diagnostics, early stage clinical trials, therapeutic cell sorting, Phase 1 clinical trials, cellular immunotherapies, ABRF, USEDit, NACCSR

Availability: Open

Resource Name: Fred Hutchinson Cancer Center Flow Cytometry Core Facility

Resource ID: SCR_022613

Alternate IDs: ABRF_1533

Alternate URLs: https://coremarketplace.org/RRID:SCR_022613/?citation=1

Record Creation Time: 20220802T050144+0000

Record Last Update: 20260819T044551+0000

Ratings and Alerts

No rating or validation information has been found for Fred Hutchinson Cancer Center Flow Cytometry Core Facility.

No alerts have been found for Fred Hutchinson Cancer Center Flow Cytometry Core Facility.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 65 mentions in open access literature.

Listed below are recent publications. The full list is available at [NIF](#) .

Harari S, et al. (2026) Mutations to the HCoV-229E spike have counterbalancing effects on serum antibody neutralization and receptor binding. bioRxiv : the preprint server for biology.

Li C, et al. (2026) In vivo HSC gene therapy enables sustained eCD4-Ig expression for SIV prevention. Molecular therapy. Advances, 34(1), 201683.

Zambidis AE, et al. (2026) CaptureBody - an anti-CD45 × anti-IgG bispecific antibody enables accurate unmixing for spectral flow cytometry. bioRxiv : the preprint server for biology.

Hagen MW, et al. (2026) The bone marrow niche and hematopoietic system are distinctly remodeled by CD45-targeted astatine-211 radioimmunotherapy. Blood advances, 10(7), 2452.

Taber A, et al. (2026) Human lung ?? T cells maintain functionality during inflammatory lung disease. bioRxiv : the preprint server for biology.

Yu TC, et al. (2026) Pleiotropic mutational effects on function and stability constrain the antigenic evolution of influenza haemagglutinin. Nature ecology & evolution, 10(3), 452.

Zhang S, et al. (2026) A CD8?? co-receptor modified to contain an intracellular CD28 signaling tail enhances TCR-engineered T cell function independent of solid-tumor-associated co-stimulatory ligands. Nature communications, 17(1), 753.

Reed SJ, et al. (2026) Junction opener enables CAR T cell treatment of solid tumors. Scientific reports, 16(1).

Chhan CB, et al. (2026) Transgenic mouse-derived human monoclonal antibodies targeting EBV gp350 and gp42 provide basis for therapeutic development. Cell reports. Medicine, 7(2), 102618.

Konecny AJ, et al. (2026) An at-home blood collection device for remote immune monitoring by high-parameter flow cytometry. JCI insight, 11(7).

Hamm DC, et al. (2026) KLF18 is a necessary component of the DUX4-initiated transcriptional network and a candidate locus for phenotypic diversity. Genes & development, 40(11-12), 873.

Murray J, et al. (2026) Engraftment of gene-edited hematopoietic stem cells after antibody-drug conjugate conditioning in nonhuman primates. Blood advances, 10(4), 1094.

Ahn JJ, et al. (2026) Influenza hemagglutinin subtypes have different sequence constraints despite sharing extremely similar structures. bioRxiv : the preprint server for biology.

Larsen BB, et al. (2026) Functional and antigenic constraints on the Nipah virus fusion protein. bioRxiv : the preprint server for biology.

Kuhlmann AS, et al. (2026) Comparative In Vitro Evaluation of Anti-HIV Immunotoxin, Antibody-Drug Conjugate, and Radioimmunoconjugate Targeted by the Same Antibody. Antibodies (Basel, Switzerland), 15(1).

Wan YH, et al. (2026) Monoclonal neutralizing antibodies elicited by infection with Kaposi sarcoma-associated herpesvirus reveal critical sites of vulnerability on gH/gL. PLoS pathogens, 22(1), e1013772.

Ahn JJ, et al. (2026) Influenza hemagglutinin subtypes have different sequence constraints despite sharing extremely similar structures. Virus evolution, 12(1), veag018.

Castelli JMP, et al. (2026) In vivo production of an anti-HIV antibody in mice by non-viral gene knockin in primate hematopoietic stem and progenitor cells. Molecular therapy : the journal of the American Society of Gene Therapy, 34(5), 2754.

Kuhlmann AS, et al. (2026) In vitro evaluation of anti-HIV radioimmunoconjugates labeled with astatine-211, thorium-227 and actinium-225. Nuclear medicine and biology, 154-155, 109602.

Avanessian SC, et al. (2026) IL2/IL15 Signaling Induces NK Cell Production of FLT3LG, Augmenting Anti-PD-1 Immunotherapy. Cancer immunology research, 14(1), 122.