Resource Summary Report

Generated by NIF on Sep 5, 2026

pDD104 (Peft-3::Cre)

RRID:Addgene_47551
Type: Plasmid

Proper Citation

RRID:Addgene_47551

Plasmid Information

URL: http://www.addgene.org/47551

Proper Citation: RRID:Addgene_47551

Insert Name: Cre Recombinase

Bacterial Resistance: Ampicillin

Defining Citation: PMID:23995389

Vector Backbone Description: Backbone Size:9539; Vector Backbone:pCFJ212; Vector Types:Worm Expression, Cre/Lox; Bacterial Resistance:Ampicillin

Comments: For more information on Goldstein Lab CRISPR Plasmids please refer to: http://www.addgene.org/crispr/Goldstein/ Please note: eft-3 has officially been changed to eef-1A.1 Please see the eef-1A.1 WormBase entry for details: http://www.wormbase.org/species/c_elegans/gene/WBGene00001168#05-9g-3

Plasmid Name: pDD104 (Peft-3::Cre)

Record Creation Time: 20220422T222245+0000

Record Last Update: 20260905T075713+0000

Ratings and Alerts

No rating or validation information has been found for pDD104 (Peft-3::Cre).

No alerts have been found for pDD104 (Peft-3::Cre).

Data and Source Information

Source: Addgene

Usage and Citation Metrics

We found 17 mentions in open access literature.

Listed below are recent publications. The full list is available at NIF .

Park S, et al. (2024) Dopey-dependent regulation of extracellular vesicles maintains neuronal morphology. bioRxiv : the preprint server for biology.

Park S, et al. (2024) Dopey-dependent regulation of extracellular vesicles maintains neuronal morphology. Current biology : CB, 34(21), 4920.

Kurashina M, et al. (2023) Targeting endogenous proteins for spatial and temporal knockdown using auxin-inducible degron in Caenorhabditis elegans. STAR protocols, 4(1), 102028.

Marom R, et al. (2023) Dominant negative variants in KIF5B cause osteogenesis imperfecta via down regulation of mTOR signaling. PLoS genetics, 19(11), e1011005.

Hendi A, et al. (2022) Channel-independent function of UNC-9/Innexin in spatial arrangement of GABAergic synapses in C. elegans. eLife, 11.

Negishi T, et al. (2022) The auxin-inducible degron 2 (AID2) system enables controlled protein knockdown during embryogenesis and development in Caenorhabditis elegans. Genetics, 220(2).

Moseley-Alldredge M, et al. (2022) A role for the Erk MAPK pathway in modulating SAX-7/L1CAM-dependent locomotion in Caenorhabditis elegans. Genetics, 220(2).

Kurashina M, et al. (2021) Sustained expression of unc-4 homeobox gene and unc-37/Groucho in postmitotic neurons specifies the spatial organization of the cholinergic synapses in C. elegans. eLife, 10.

Lee IH, et al. (2021) Stress-Induced Neural Plasticity Mediated by Glial GPCR REMO-1 Promotes C.??elegans Adaptive Behavior. Cell reports, 34(2), 108607.

Hsu TY, et al. (2021) C. elegans orthologs MUT-7/CeWRN-1 of Werner syndrome protein regulate neuronal plasticity. eLife, 10.

Manage KI, et al. (2020) A tudor domain protein, SIMR-1, promotes siRNA production at piRNA-targeted mRNAs in C. elegans. eLife, 9.

Lu M, et al. (2019) Gradient-independent Wnt signaling instructs asymmetric neurite pruning in C. elegans. eLife, 8.

Uebel CJ, et al. (2018) Distinct regions of the intrinsically disordered protein MUT-16 mediate assembly of a small RNA amplification complex and promote phase separation of Mutator foci. PLoS genetics, 14(7), e1007542.

Schwartz ML, et al. (2016) SapTrap, a Toolkit for High-Throughput CRISPR/Cas9 Gene Modification in Caenorhabditis elegans. Genetics, 202(4), 1277.

Huelgas-Morales G, et al. (2016) The Stress Granule RNA-Binding Protein TIAR-1 Protects Female Germ Cells from Heat Shock in Caenorhabditis elegans. G3 (Bethesda, Md.), 6(4), 1031.

Bone CR, et al. (2016) Nuclei migrate through constricted spaces using microtubule motors and actin networks in C. elegans hypodermal cells. Development (Cambridge, England), 143(22), 4193.

Calarco JA, et al. (2015) Creating Genome Modifications in C. elegans Using the CRISPR/Cas9 System. Methods in molecular biology (Clifton, N.J.), 1327, 59.