Resource Summary Report

Generated by NIF on Jul 28, 2026

Anti-CD38 [OKT10]-APC

RRID:AB_2819277
Type: Antibody

Proper Citation

(NIH Nonhuman Primate Reagent Resource Cat# PR-3801, RRID:AB_2819277)

Antibody Information

URL: http://antibodyregistry.org/AB_2819277

Proper Citation: (NIH Nonhuman Primate Reagent Resource Cat# PR-3801, RRID:AB_2819277)

Target Antigen: CD38

Clonality: monoclonal

Comments: Originating vendor of this resource; Applications: flow cytometry Info: APC-conjugate of OKT10 for immunophenotyping macaques.

Antibody Name: Anti-CD38 [OKT10]-APC

Description: This monoclonal targets CD38

Target Organism: rhesus

Clone ID: clone OKT10

Antibody ID: AB_2819277

Vendor: NIH Nonhuman Primate Reagent Resource

Catalog Number: PR-3801

Record Creation Time: 20260218T234914+0000

Record Last Update: 20260226T001125+0000

Ratings and Alerts

No rating or validation information has been found for Anti-CD38 [OKT10]-APC.

No alerts have been found for Anti-CD38 [OKT10]-APC.

Data and Source Information

Source: Antibody Registry

Usage and Citation Metrics

We found 21 mentions in open access literature.

Listed below are recent publications. The full list is available at NIF .

Ma KM, et al. (2025) HIV broadly neutralizing antibody precursors to the Apex epitope induced in nonhuman primates. Science immunology, 10(110), eadt6660.

Schiffner T, et al. (2024) Vaccination induces broadly neutralizing antibody precursors to HIV gp41. Nature immunology, 25(6), 1073.

Steichen JM, et al. (2024) Vaccine priming of rare HIV broadly neutralizing antibody precursors in nonhuman primates. Science (New York, N.Y.), 384(6697), eadj8321.

Araki D, et al. (2024) cMPL-Based Purification and Depletion of Human Hematopoietic Stem Cells: Implications for Pre-Transplant Conditioning. bioRxiv : the preprint server for biology.

Lee JH, et al. (2022) Long-primed germinal centres with enduring affinity maturation and clonal migration. Nature, 609(7929), 998.

Esaulova E, et al. (2021) The immune landscape in tuberculosis reveals populations linked to disease and latency. Cell host & microbe, 29(2), 165.

Acharya A, et al. (2021) Chronic morphine administration differentially modulates viral reservoirs in SIVmac251 infected rhesus macaque model. Journal of virology, 95(5).

Bricker KM, et al. (2020) Therapeutic vaccination of SIV-infected, ART-treated infant rhesus macaques using Ad48/MVA in combination with TLR-7 stimulation. PLoS pathogens, 16(10), e1008954.

Chaudhary O, et al. (2018) Inhibition of p38 MAPK in combination with ART reduces SIV-induced immune activation and provides additional protection from immune system deterioration. PLoS pathogens, 14(8), e1007268.

Gori JL, et al. (2017) Endothelial Cells Promote Expansion of Long-Term Engrafting Marrow Hematopoietic Stem and Progenitor Cells in Primates. Stem cells translational medicine, 6(3), 864.

Jaworski JP, et al. (2017) Pre-existing neutralizing antibody mitigates B cell dysregulation and enhances the Env-specific antibody response in SHIV-infected rhesus macaques. PloS one, 12(2), e0172524.

Martinez-Murillo P, et al. (2016) CD138 and CD31 Double-Positive Cells Comprise the Functional Antibody-Secreting Plasma Cell Compartment in Primate Bone Marrow. Frontiers in immunology, 7, 242.

Neumann B, et al. (2016) Comprehensive panel of cross-reacting monoclonal antibodies for analysis of different immune cells and their distribution in the common marmoset (Callithrix jacchus). Journal of medical primatology, 45(3), 139.

Gori JL, et al. (2015) Vascular niche promotes hematopoietic multipotent progenitor formation from pluripotent stem cells. The Journal of clinical investigation, 125(3), 1243.

Neumann B, et al. (2015) Characterization of B and plasma cells in blood, bone marrow, and secondary lymphoid organs of rhesus macaques by multicolor flow cytometry. Journal of leukocyte biology, 97(1), 19.

Estes JD, et al. (2015) Antifibrotic therapy in simian immunodeficiency virus infection preserves CD4+ T-cell populations and improves immune reconstitution with antiretroviral therapy. The Journal of infectious diseases, 211(5), 744.

Hao XP, et al. (2015) Experimental colitis in SIV-uninfected rhesus macaques recapitulates important features of pathogenic SIV infection. Nature communications, 6, 8020.

Neumann B, et al. (2015) OMIP-026: Phenotypic analysis of B and plasma cells in rhesus macaques. Cytometry. Part A : the journal of the International Society for Analytical Cytology, 87(9), 800.

Tabb B, et al. (2013) Reduced inflammation and lymphoid tissue immunopathology in rhesus macaques receiving anti-tumor necrosis factor treatment during primary simian immunodeficiency virus infection. The Journal of infectious diseases, 207(6), 880.

Del Prete GQ, et al. (2012) Restricted replication of xenotropic murine leukemia virus-related virus in pigtailed macaques. Journal of virology, 86(6), 3152.