

Resource Summary Report

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CD279

RRID:AB_11153482

Type: Antibody

Proper Citation

BD Biosciences Cat# 562516, RRID:AB_11153482

Antibody Information

URL: http://antibodyregistry.org/AB_11153482

Proper Citation: BD Biosciences Cat# 562516, RRID:AB_11153482

Target Antigen: CD279 (PD-1)

Host Organism: mouse

Clonality: monoclonal

Comments: Applications: Flow cytometry

Antibody Name: CD279

Description: This monoclonal targets CD279 (PD-1)

Target Organism: human

Antibody ID: AB_11153482

Vendor: BD Biosciences

Catalog Number: 562516

Record Creation Time: 20250819T215423+0000

Record Last Update: 20250820T002847+0000

Ratings and Alerts

No rating or validation information has been found for CD279.

No alerts have been found for CD279.

Data and Source Information

Source: [Antibody Registry](#)

Usage and Citation Metrics

We found 17 mentions in open access literature.

Listed below are recent publications. The full list is available at [NIF](#) .

Gu S, et al. (2025) Transient regulatory T cell manipulation is limited by anti-antibody responses in HIV-1 envelope immunized rhesus macaques. *iScience*, 28(8), 113191.

Long GV, et al. (2025) Neoadjuvant triplet immune checkpoint blockade in newly diagnosed glioblastoma. *Nature medicine*, 31(5), 1557.

Wang D, et al. (2025) Allogeneic CD19-targeted CAR-T therapy in refractory systemic lupus erythematosus achieved durable remission. *Med (New York, N.Y.)*, 100749.

Wang K, et al. (2024) Combination anti-PD-1 and anti-CTLA-4 therapy generates waves of clonal responses that include progenitor-exhausted CD8+ T cells. *Cancer cell*, 42(9), 1582.

Wang X, et al. (2024) Allogeneic CD19-targeted CAR-T therapy in patients with severe myositis and systemic sclerosis. *Cell*, 187(18), 4890.

Veith I, et al. (2024) Assessing personalized responses to anti-PD-1 treatment using patient-derived lung tumor-on-chip. *Cell reports. Medicine*, 5(5), 101549.

Mthembu M, et al. (2024) Dysfunctional bronchoalveolar effector memory CD8+ T cells in tuberculosis-exposed people living with antiretroviral-naïve HIV infection. *iScience*, 27(11), 111137.

Trinh T, et al. (2023) CX3CR1 deficiency-induced TIL tumor restriction as a novel addition for CAR-T design in solid malignancies. *iScience*, 26(4), 106443.

Liu H, et al. (2022) Optimal target saturation of ligand-blocking anti-GITR antibody IBI37G5 dictates FcγR-independent GITR agonism and antitumor activity. *Cell reports. Medicine*, 3(6), 100660.

Rasmussen TA, et al. (2022) Memory CD4+ T cells that co-express PD1 and CTLA4 have reduced response to activating stimuli facilitating HIV latency. *Cell reports. Medicine*, 3(10), 100766.

Ducoin K, et al. (2022) Defining the Immune Checkpoint Landscape in Human Colorectal Cancer Highlights the Relevance of the TIGIT/CD155 Axis for Optimizing Immunotherapy. *Cancers*, 14(17).

Law H, et al. (2022) Early expansion of CD38+ICOS+ GC Tfh in draining lymph nodes during influenza vaccination immune response. *iScience*, 25(1), 103656.

Herati RS, et al. (2021) Vaccine-induced ICOS+CD38+ circulating Tfh are sensitive biosensors of age-related changes in inflammatory pathways. *Cell reports. Medicine*, 2(5), 100262.

Procaccini C, et al. (2021) Signals of pseudo-starvation unveil the amino acid transporter SLC7A11 as key determinant in the control of Treg cell proliferative potential. *Immunity*, 54(7), 1543.

Abdul-Jawad S, et al. (2021) Acute Immune Signatures and Their Legacies in Severe Acute Respiratory Syndrome Coronavirus-2 Infected Cancer Patients. *Cancer cell*, 39(2), 257.

Singh A, et al. (2020) Innate Lymphoid Cell Activation and Sustained Depletion in Blood and Tissue of Children Infected with HIV from Birth Despite Antiretroviral Therapy. *Cell reports*, 32(11), 108153.

Koguchi Y, et al. (2016) A Semi-automated Approach to Preparing Antibody Cocktails for Immunophenotypic Analysis of Human Peripheral Blood. *Journal of visualized experiments : JoVE*(108), e53485.